

# Kazu Buffer Feedback Form

**Name:** Dong-Yeon Lee

**Lab:** (Brutnell lab)

**Material:** Setaria leaf

**Primer set:** Gene specific and pZmUni-HPT

**Amplification size (kb):** 0.3kb and .6kb

**PCR taq:** Dream Taq

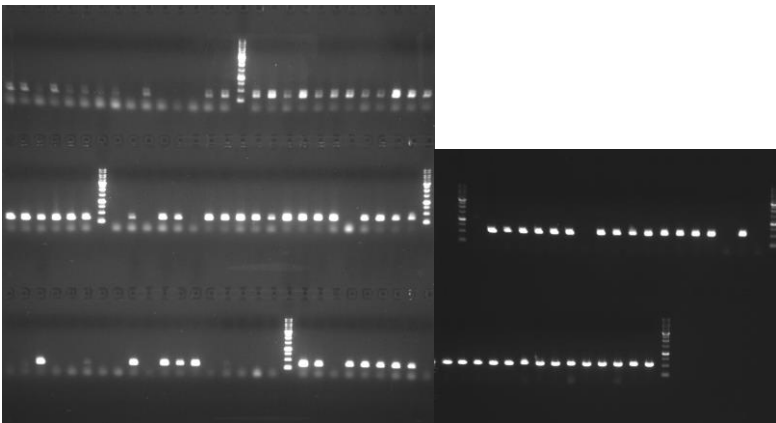
## PCR amplification:

Reagent	Volume
Kazu DNA extract	2 $\mu$ l
Taq	0.2 $\mu$ l
Forward primer	0.5 $\mu$ l
Reverse primer	0.5 $\mu$ l
10x Taq Buffer	2 $\mu$ l
dNTP(10mM)	0.4 $\mu$ l
H <sub>2</sub> O	14.4 $\mu$ l
Total volume	20 $\mu$ l

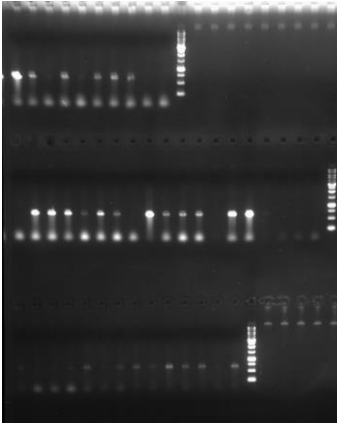
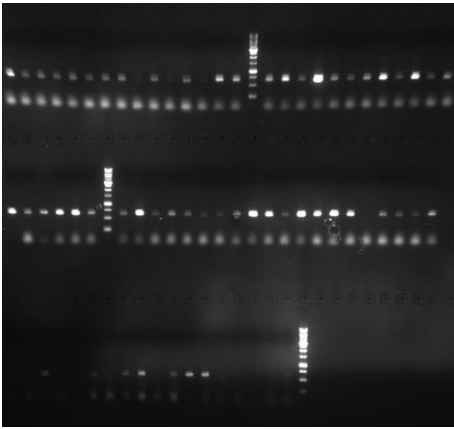
Step	Temperature	Time	Cycles
Initial			
Denaturation	95°C	2 min	N/A
Denaturation	95°C	10 sec	40
Annealing	58°C	10 sec	
Extension	72°C	1 min	
Final Extension	72°C	5 min	N/A
Hold	12 °C	10 min	N/A

## PCR Gel Picture:

<gene specific>



<pZmUbi-HPT>



Would you prefer using Kazu Buffer over your current lab DNA extraction protocol?  Yes  No

Why or why not?

It depends on purpose of DNA usage.

Other Comments:

Feel like still need to improve some procedures for high throughput analysis for consistent results.

Please keep polishing standardization and make big improvement.